6-14-22

Just some tips and tricks for you before we actually revise the confusing protocol…

1. Try to use multi-channel pipette as much as possible when plating any liquid onto plates
2. **PPO and PER use clear plates, beta-glucosidase uses black plates**
3. If using multi-channel pipettes and there is only a little volume for your solution, use the large plastic beakers so the pipette can fit in. Double or triple volume amount if the liquid isn’t enough
   1. Only the 400mL beakers will work for multi-channel pipettes. Don’t use anything smaller…
   2. If reagent is light-sensitive just wrap some aluminum foil around the beaker
4. To make 0.3% H2O2 solution from 3% stock solution, combine 1 mL stock solution with 9 mL of milli-Q water
5. **Everything needs to be autoclaved if using for sample holding. Otherwise, ethanol spatulas, etc. Even water, only use milli-Q for everything.**
6. When sucking up solution from pipette tips quickly check to make sure each tip has roughly the same amount of liquid in it, otherwise push tips in more and try again
7. Double and triple check that your desired volume matches the volume you’re sucking up with your pipette. The volumes switch a lot throughout the protocol.
8. You’re going to need a lot more than 1L of sodium acetate buffer most likely. Remember each sample needs 125 mL and each plate needs roughly 6-7 mL. Better to have a little more to get you through the week.
9. When making L-DOPA solution, you can add both L-DOPA and EDTA into 100 mL total milli-Q water. Make sure the water is a little heated beforehand in order to fully dissolve L-DOPA (~1 minute in microwave).
10. Remember to dilute MUB solution every day, don’t use stock for plating.
11. Make sure to label plates with which enzyme used as well as samples/sample order
12. Mix samples with sodium acetate by using blender thing for 1-minue on high (set timer), then transfer to plate to stir samples (with sterile stir bar).
13. Check Jenni’s Protocol on how to store solutions located at the table at the end. It’s pretty fire.
14. **Make sure to record weights of each sample used prior to discarding beaker contents**
15. Autoclave all dirty graduated cylinders and beakers used the day before. There is a limited number of beakers that can be used for multichannel pipettors (~20).